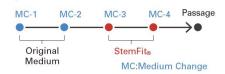
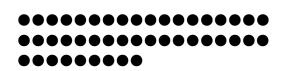
- Tips on transitioning cells to StemFit® medium
  - •Switch culture medium to StemFit<sub>®</sub> 2 3 days prior to passage
  - <Example>



•Seed the cells at a higher density (>1.0 x 10<sup>5</sup> cells per well (6-well plate))





## Feeder free medium for ES/iPS cells



Key points for successful single-cell passage



Robust and reproducible culture

Quantitative culture



**High fold expansion** 

 $\sim$ 100X expansion / passage

## Single-cell passage brief protocol example (6-well plate) and tips



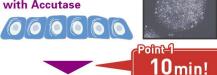
Aspirate the medium and wash once with 2 mL of PBS



Add 500 μl/well of Accutase and incubate at 37 °C for 10 min

- \* TrypLE™ can also be used for cell dissociation
- \* Incubation time may vary depending on the matrix





 Gaps in the colonies appear and dissociation of colonies is apparent



Whole colony can be smoothly detached

Gently pipette the cells to fully dissociate and transfer cells to a 15 ml tube filled with 500 µl of culture medium containing 10 µM Y-27632



Count the cells and centrifuge the tubes

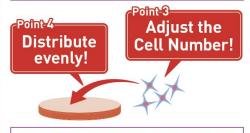


Aspirate the medium and resuspend cells with culture medium containing 10 µM Y-27632



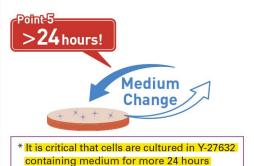
Add 10-20 μl (1.0-2.0 x 10<sup>4</sup> cells) of resuspended cells per well in 1.5 mL of culture medium containing 10 μM Y-27632

- \* It is important to adjust the plating cell number for different lines of hPSCs
- \* Try higher seeding density when cell or colony quantity is insufficient (See also Tips on transitioning cells to StemFit<sub>®</sub> medium)



\* Immediately distribute the cells evenly over the plate surface to avoid uneven attachment

After >24 hours of culture, replace with fresh culture medium without Y-27632



7 Perform medium change

